

Pig post-mortem guide

Equipment required:

- standard post-mortem kit including boning and skinning knives, scissors, scalpel blades, forceps
- large shears or pruning secateurs to cut through ribs and jaw, bone saw
- shears or small axe and hammer for brain removal
- dry swabs, swabs in media (viral transport media, Amies media/charcoal), specimen jars for individual fresh samples, 10% formalin, large containers for pooled formalin fixed samples
- personal protection equipment – overalls, gloves, mask, eye protection
- laboratory submission forms, permanent markers.

Sampling guidelines:

- The base sample list is not exhaustive – consider submitting any other samples that may be useful (i.e. feed, water etc) and any lesions or abnormalities.
- Fresh and fixed samples of any discrete **lesions** should also be submitted.
- Consider sampling cohort animals (i.e. blood, faeces, swabs).
- Complete a **submission form** to submit with samples. Tape to the **outside** of the esky/box of samples. Submissions forms and detailed packaging instructions are available on agric.wa.gov.au.
- Consider sending photographs or videos to the lab at DDL@dpird.wa.gov.au

Base sample list – pigs

Ante-mortem samples	Post-mortem samples	
	Fresh	Fixed
Blood: Lithium heparin Clotted EDTA (+ smears) Faeces Swabs of discharges etc	Spleen Lymph nodes Tonsil Ileum Brain (swab or small section of cortex) Liver Lung Kidney Stomach contents Faeces	Brain (submitted whole) Kidney Liver Lung Spleen Lymph nodes Tonsil Stomach Small and large intestine Ileum Heart Skeletal muscle
Chill bloods; consider spinning and separating serum and clot for transport. Do not send sharps. For swabs use the appropriate media then chill: • Amies or charcoal for bacteria • viral transport media for viruses and PCR • dry swabs (as a last resort) +/- a small drop of sterile saline for PCR. Special media (e.g. Mycoplasma media) available on request.	Collect a 3x3cm piece of tissue and store in a sterile container / sample pot. Refrigerate or chill – DO NOT freeze.	Pool tissues into 10% neutral buffered formalin at a ratio of 10 parts formalin to 1 part tissue. Keep at room temperature. Seal containers and package well to avoid spills.