Pig post-mortem guide

Equipment required:
- standard post-mortem kit including boning and skinning knives, scissors, scalpel blades, forceps
- large shears or pruning secateurs to cut through ribs and jaw, bone saw
- shears or small axe and hammer for brain removal
- dry swabs, swabs in media (viral transport media, Amies media/charcoal), specimen jars for individual fresh samples, 10% formalin, large containers for pooled formalin fixed samples
- personal protection equipment – overalls, gloves, mask, eye protection
- laboratory submission forms, permanent markers.

Sampling guidelines:
- The base sample list is not exhaustive – consider submitting any other samples that may be useful (i.e. feed, water etc) and any lesions or abnormalities.
- Fresh and fixed samples of any discrete lesions should also be submitted.
- Consider sampling cohort animals (i.e. blood, faeces, swabs).
- Complete a submission form to submit with samples. Tape to the outside of the esky/box of samples. Submissions forms and detailed packaging instructions are available on agric.wa.gov.au.
- Consider sending photographs or videos to the lab at DDLS@dpiwd.wa.gov.au

Base sample list – pigs

<table>
<thead>
<tr>
<th>Ante-mortem samples</th>
<th>Post-mortem samples</th>
<th>Fixed</th>
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</thead>
<tbody>
<tr>
<td>Blood: Lithium heparin Clotted EDTA (+ smears)</td>
<td>Fresh Spleen Lymph nodes Tonsil Ileum Brain (swab or small section of cortex) Liver Lung Kidney Stomach contents Faeces</td>
<td>Fixed Brain (submitted whole) Kidney Liver Lung Spleen Lymph nodes Tonsil Stomach Small and large intestine Ileum Heart Skeletal muscle</td>
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<td>Faeces Swabs of discharges etc</td>
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Chill bloods; consider spinning and separating serum and clot for transport. Do not send sharps.
For swabs use the appropriate media then chill:
- Amies or charcoal for bacteria
- viral transport media for viruses and PCR
- dry swabs (as a last resort) +/- a small drop of sterile saline for PCR.
Special media (e.g. Mycoplasma media) available on request.

Collect a 3x3cm piece of tissue and store in a sterile container / sample pot. Refrigerate or chill – DO NOT freeze.

Pool tissues into 10% neutral buffered formalin at a ratio of 10 parts formalin to 1 part tissue.
Keep at room temperature. Seal containers and package well to avoid spills.

Contact the lab on (08) 9368 3351 prior to submission if you suspect a reportable or exotic disease.